

## IMPACT OF *TRICHODERMA ASPERELLUM* FERMENTED AGRICULTURAL WASTE-BASED FEED ON BROILER CHICKEN GROWTH AND MEAT QUALITY

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### ABSTRACT

The demand for poultry products is increasing due to the rising global population. This has led to high prices and inadequacy of traditional poultry feed ingredients, creating a need to explore alternative feed sources like agricultural wastes. The present study was conducted to assess the effect of *Trichoderma asperellum*-fermented agricultural waste-based feed on the growth and meat quality of broiler. Wheat bran and rice husk were selected as agricultural waste-based substrates for solid state fermentation (SSF). A total of 25 one-day old broiler chicks were distributed into 5 treatment groups randomly: group 1 was fed completely on basic feed mixture (control), group 2 was fed on diet with 10% unfermented wheat bran (WB), group 3 was fed on diet with 10% fermented wheat bran (FWB), group 4 was fed on diet with 10% unfermented rice husk (RH) and group 5 was fed on diet with 10% fermented rice husk (FRH). The experiment was conducted for 5 weeks. Nutrient analysis showed that solid-state fermentation (SSF) improved the protein and carbohydrate content of the substrates. Statistical analysis of growth performance showed that there was a significant ( $P < 0.05$ ) improvement in body weight in the group fed on diet with 10% fermented wheat bran (FWB) compared to control group. The group fed on 10% fermented rice husk (FRH) maintained a similar body weight as compared to control group. The groups fed on 10% unfermented wheat bran (WB) and 10% unfermented rice husk (RH) had lower body weight than control group. No significant changes were observed in meat quality parameters among all the treatment groups. In conclusion, fermenting the wheat barn through fungus *Trichoderma asperellum* not only improved the nutrient content by degrading complex components but also had a positive effect on chicken growth. This study suggests that wheat bran fermented by *Trichoderma asperellum* can be added to broiler diets at up to 10% without any deleterious effect on growth and meat quality.

## 1. INTRODUCTION

The poultry industry is experiencing feed price fluctuations due to rising costs of key feed ingredients prompting poultry nutritionists to explore alternative feed sources (Sugiharto et al., 2017). Some developing nations are producing various alternative ingredients such as wheat bran, rice bran, cottonseed meal, palm kernel cake and copra meal derived from agricultural wastes and byproducts. These feedstuffs are more affordable compared to main grain crops. However, their high fibre contents limit their usefulness for monogastric animals (Oguri et al., 2013; Chu et al., 2017).

For example, wheat bran and rice husk are among such ingredients with restricted applications in monogastric diets. The outer coat of seed from flour mills is known as wheat bran (Saini et al., 2023). It has the potential to replace yellow corn in poultry diets partially or completely (Alshelmani et al., 2021). WB includes a significant quantity of protein (160 g/kg), complete dietary fibre (451 g/kg), carbohydrates (177 g/kg), minerals (61.5 g/kg) and fat (47g/kg) (Teng et al., 2017; Junejo et al., 2019). It has high fibre content but low metabolizable energy (ME), thus its use in poultry diets is limited. Its nutrient content can be improved by microbial fermentation. It has been found that replacement of 10% basal diet with fermented wheat

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bran can be incorporated to broiler diets without any deleterious effect (Chu et al., 2017).

Rice husk is an agricultural waste generated during rice grain milling. Currently, large piles of rice husks continue to accumulate in many milling areas, where they are often burned for heat or discarded as waste. This disposal method poses significant environmental risks, including pollution and resource wastage. The recent trend in livestock farming is to utilize agricultural waste as substitute feed ingredients in broiler and other animal feeding trials. Although rice husk has long been recognized as a potential feed ingredient, its high silica content, high fiber levels, and harsh texture make it difficult to degrade, limiting its application in animal feed (Ikpe and Oko, 2019). Fermentation is a technique that enhances nutritional value of feed ingredients by reducing crude fiber and antinutritional contents (Predescu et al., 2024). To improve the nutrient content of agricultural wastes and by-products, researchers have been using fungal inoculums in recent years (Sharma and Arora, 2011; Choudhary et al., 2016). Recent studies have employed various *Trichoderma* species to ferment wheat bran and rice husk, breaking down complex compounds such as cellulose, hemicellulose, lignin, and non-starch polysaccharides into simpler forms. This process enhances their digestibility for broilers and improves their nutrient content (Aderolu et al., 2007; Chu et al., 2017).

*Trichoderma* are free living and filamentous fungi. They are usually found on decomposing plants, wood, and in the plant rhizosphere, where they provoke general tolerance against pathogens (Afzal et al., 2021; Jangir et al., 2017). Species of *Trichoderma* are good manufacturers of cellulases. They have strong penetration ability into non-soluble substrates and are therefore considered highly suitable for fermenting materials containing lignin and cellulose (Elmasry et al., 2017). Chiang et al. (2009) described that solid state fermentation enhanced the digestibility of nutrients, increased lactobacilli count in the large intestine, improved the gut morphology of broiler chickens and reduced the cost of poultry farming. Alshelmani et al. (2017) stated that broilers fed on feed compositions accompanied by *Paenibacillus polymyxa*-fermented palm kernel cake exhibited improved growth rates and meat quality. Likewise, Chu et al. (2017) pointed out that *Trichoderma pseudokoningii*-fermented wheat bran supplemented diets improved the growth rate and intestinal morphology of broilers. Similarly, Gungor and Erener (2020) showed that broilers given a diet supplemented with 1% *Aspergillus niger*-fermented sour cherry kernel improved weight gain and feed conversion ratio

in broilers. (Aderolu et al., 2007) indicated that *Trichoderma viride* fermented rice husk showed improved degradation of cellulosic content and increased energy content. Elmasry et al. (2017) indicated that wheat bran fermented with *Trichoderma longibrachiatum* as 10% feed additive improved the growth performance and feed consumption of broilers than control group. According to Hatta et al. (2014) broilers fed diets containing copra meal fermented with *Trichoderma viride* exhibited similar growth performance to the control group. Similarly, Marcinčák et al. (2018) described that corn meal fermented with *Umbelopsis isabellina* improved the growth and quality of meat of broilers fed on it as compared to control group.

Only a few microbes have been investigated so far to improve the nutritional value and digestibility of agricultural waste-based feed ingredients. In the current study, the potential of a lignocellulose degrading fungus *Trichoderma asperellum* was evaluated to improve the nutritive value of agro-waste products and its effects on broiler growth and meat quality were assessed.

## 2. MATERIALS AND METHODS

### *Collection and culturing of fungus*

The previously isolated and identified fungus, *Trichoderma asperellum*, was procured from the Department of Biology, Lahore Garrison University. Potato dextrose agar (PDA) was utilized as the growth medium. The media was prepared and poured into petri plates under sterile conditions and allowed to solidify. The fungal inoculum was inoculated onto PDA plates with the help of sterile inoculating loop and incubated at 25-30°C for 7 days.

### *Collection of substrate samples*

Wheat bran and rice husk were selected as substrates for solid state fermentation. Both were purchased from the local market and stored under ambient conditions until fermentation.

### *Pretreatment, Solid state fermentation (SSF), and Nutrient analysis*

Aluminum boxes each containing 100 grams of wheat bran and rice husk were filled with deionized water to achieve 50% moisture content, mixed thoroughly, packed, and autoclaved. The boxes were then cooled to room temperature (Chu et al., 2017). For solid-state fermentation, a sterile saline solution was added to cultured PDA plates, and fungal spores of *Trichoderma asperellum* were harvested and suspended at a concentration of  $1 \times 10^5$  to  $1 \times 10^7$  spores per ml. Each kilogram of wheat bran and rice husk was

inoculated with 200 ml of spore suspension, packed, and incubated at 30°C for two weeks.

Post-incubation, the samples were autoclaved, washed, sun-dried, and stored at room temperature (Elmasry et al., 2017). For nutrient analysis, the fermented and unfermented samples were mixed with phosphate buffer, ground, and centrifuged. The supernatants were collected for protein and carbohydrate quantification. Protein content was measured using the Lowry method, with absorbance recorded at 660 nm (Satpathy et al., 2020), while carbohydrates content was determined using the Anthrone test, with absorbance recorded at 630 nm. Results were expressed as milligrams of protein or carbohydrate per gram of substrate.

#### **Experimental birds, Housing, and Categorization**

Twenty-five healthy one-day broiler chicks were separated from the flock and housed in a local animal shed in Gujranwala city. The chicks were weighed and randomly assigned to five dietary groups: Group 1 was fed a basic feed mixture (control), Group 2 received a diet with 10% unfermented wheat bran (WB), Group 3 received a diet with 10% fermented wheat bran (FWB), Group 4 was fed a diet with 10% unfermented rice husk (RH) and Group 5 received a diet with 10% fermented rice husk (FRH). The experiment was conducted over 35 days, during which the chicks had ad libitum access to feed and water.

#### **Statistical analysis**

The data for total protein and carbohydrate content was subjected to two sample T-test, assuming equal variances, in SPSS software to evaluate the statistical effect of microbial fermentation on wheat bran and rice husk. Data on body weight and meat quality were subjected to one-way ANOVA using SPSS software to assess the effects of experimental diets on the treatment groups. Significant differences between treatment groups were considered at a significance level of  $P < 0.05$ .

### **3. RESULTS AND DISCUSSION**

#### **Protein analysis**

The protein content of unfermented and *Trichoderma asperellum*-fermented wheat bran and rice husk was analyzed through Lowry method, with results presented in Table 1. Fermentation with *T. asperellum* significantly increased the protein content of wheat bran from 19.06±0.13 mg/g to 54.51±0.09 mg/g, and rice husk from 10.56±0.06 mg/g to 10.80±0.06 mg/g. According to the t-test analysis, the increase in protein concentration for both fermented wheat bran and fermented rice husk compared to their unfermented

counterparts was statistically significant ( $P < 0.05$ ), as shown in Table 1.

#### **Carbohydrate analysis**

The carbohydrate content of unfermented and *Trichoderma asperellum*-fermented wheat bran and rice husk was analyzed through anthrone test, with results presented in Table 2. Solid-state fermentation with *Trichoderma asperellum* increased the carbohydrate content of wheat bran from 16.45±0.07 mg/g to 22.58±0.14 mg/g and rice husk increased from 6.45±0.06 mg/g to 6.50±0.12 mg/g. According to t-test analysis, the increase in carbohydrate concentration for fermented wheat bran compared to unfermented wheat bran was statistically significant ( $P < 0.05$ ). However, for rice husk, the increase in carbohydrate concentration was not statistically significant ( $P > 0.05$ ), as shown in Table 2.

#### **Effect of designed feed mixtures on the growth performance**

The effects of the designed feed mixtures on the body weight of broiler chickens are presented in Table 3. One-way ANOVA revealed statistically significant differences in the body weights of chicks among the treatment groups at weeks 2, 3, 4, and 5 ( $P < 0.05$ ). A least significant difference (LSD) post hoc test showed that the group fed a diet containing standard feed with 10% fermented wheat bran (FWB) exhibited significantly higher body weight compared to the control and other treatment groups after five weeks. This result highlights the positive effect of fermented wheat bran. Figure 1 provides a graphical representation of the effects of the designed feed mixtures on the body weight of broiler chicks from week 1 to week 5.

#### **Effect of designed feed mixtures on the meat quality**

The parameters used to measure meat quality were pH and colour with results presented in Table 4. The L\*, a\*, and b\* values were used as indicators of breast meat colour. Chicken meat colour primarily depends on lightness values. One-way ANOVA revealed no significant differences in breast meat colour and pH among the treatment groups, indicating that the feed supplements did not result in any significant changes in meat quality. Figures 2 and 3 provide graphical representations of the pH and color differences between the treatment groups. The current study assessed the effect of fungal fermented agricultural waste-based feed on broiler chicken growth and meat quality, using the fungus *Trichoderma asperellum* (previously isolated and identified). Fermentation of wheat bran with *T. asperellum* increased protein content from 19.06 mg/g to 54.51 mg/g, and rice husk

protein content increased from 10.56 mg/g to 10.80 mg/g. Fungi grow on substrates and secrete various cellulose-degrading enzymes that convert non-starch polysaccharides into simple sugars. These sugars are absorbed by the fungus to stimulate metabolic activities and biosynthesis, leading to an increase in protein content (Chu et al., 2017). Furthermore, the fungi's ability to produce enzymes capable of hydrolyzing starch and non-starch polysaccharides into monosaccharides contributes to increased protein proportion, as monosaccharides are easily converted into protein (Bayitse et al., 2015). Carbohydrate content (simple sugars) of wheat bran increased from 16.45 mg/g to 22.58 mg/g after solid-state fermentation (SSF) with *T. asperellum*, while rice husk only showed a minor increase from 6.45 mg/g to 6.50 mg/g. The enzymatic activity during SSF efficiently breaks down lignocellulosic material (crude fiber) into reducing sugars, which are more digestible for poultry than crude fiber (Lin et al., 2018).

These findings align with previous studies, such as Chu et al. (2017), who reported that *T. pseudokoningii*-fermented wheat bran increased by reducing sugars and crude protein content. Similarly, Omwango et al. (2013) observed that pineapple waste fermented with *T. viride* enhanced crude protein content while reducing crude fiber. Ezekiel and Aworh (2013) showed that *T. viride*-fermented cassava peel significantly improved crude protein content while reducing crude fiber. Aderolu et al. (2007) demonstrated that *T. viride*-fermented rice husk reduced NDF and ADF (crude fiber) content. SSF was employed to ferment wheat bran and rice husk, chosen based on substrate characteristics. SSF provides better yield and simplifies downstream processing compared to submerged fermentation (Sugiharto and Ranjitkar, 2019). Cellulose, starch, and lignin are commonly used in SSF as carbon sources, with bacteria and yeast typically growing on the substrate surface, while fungi invade the substrate through mycelium (Srivastava et al., 2019).

In this study, the treatment group fed a diet with 10% fermented wheat bran (FWB) showed a significant increase in body weight (BW) after 5 weeks, reaching 1380 g/bird, compared to the control group (1304 g/bird) and the group fed 10% unfermented wheat bran (WB) (1296.2 g/bird). These results align with Elmasry et al. (2017), who reported a significant increase in BW in broilers fed on wheat bran fermented with *T. longibrachiatum* (SF1). Gungor and Erener (2020) also found improved growth in broilers fed sour cherry kernel fermented with *Aspergillus niger*. However, Hatta et al. (2014) reported that the growth rate of broilers fed fermented wheat bran was

like the control group. Similarly, Chuang et al. (2019) found no significant change in the growth of broilers fed a diet with *Aspergillus oryzae* and *Saccharomyces cerevisiae*-fermented wheat bran.

In contrast, the treatment groups fed 10% rice husk (RH) and 10% fermented rice husk (FRH) did not show significant changes in body weight compared to the control group. The group fed 10% rice husk (RH) had a slightly lower body weight (1293.4 g/bird) than the control group (1304 g/bird), but this difference was not statistically significant. The group fed 10% fermented rice husk (FRH) had a body weight (1311 g/bird) like the control group (1304 g/bird). These results are consistent with Ikiye and Oko (2019), who reported no significant change in body weight in groups fed rice husk fermented with molasses syrup. Post-slaughter, muscle pH is an important factor influencing meat quality traits, such as color, water holding capacity, and tenderness. The normal pH for breast muscle is above 5.70 (Alshelmani et al., 2016). In the present study, no significant differences were observed in pH between treatment groups (10% WB, 10% FWB, 10% RH, and 10% FRH) and the control group. However, the pH of the 10% rice husk group was slightly lower than the control group. The pH range was from 5.58 to 5.82 across all treatment groups. These findings are in line with Alshelmani et al. (2016), who reported no significant difference in breast muscle pH when palm kernel cake was treated with *Paenibacillus polymyxa* (ATCC 842). Meat color can be influenced by various factors, including nutrition, sex, age, strain, processing methods, intramuscular fat, and pH. A notable relationship exists between pH and meat color, where a decrease in pH can lead to protein denaturation, resulting in pale color. Lightness ( $L^*$ ) values are commonly used to assess meat color. In the present study, breast meat color across all treatment groups fell within the normal range ( $56 \leq L^* \leq 62$ ), indicating no significant differences in color among the groups. These results are consistent with Ashayerizadeh et al. (2018), who reported no significant effect on breast meat color and pH in broilers fed rapeseed meal fermented with *Aspergillus niger*, *Lactobacillus acidophilus*, and *Bacillus subtilis*.

#### 4. CONCLUSION

It was concluded that *Trichoderma asperellum*-fermented wheat bran and rice husk can serve as effective feed ingredients in poultry diets. *Trichoderma asperellum*-fermented wheat bran can be included in broiler diets up to 10% without negatively affecting growth or meat quality.

## 5. CONFLICT OF INTEREST

All authors declare that they have no conflict of interest regarding the publication of this article.

## REFERENCES

- Aderolu, A. Z., Iyayi, E. A., & Onilude, A. A. (2007). Changes in Nutritional Value of Rice Husk during *Trichoderma viride* Degradation. *Bulgarian Journal of Agricultural Science*, 13, 583–589.
- Afzal, I., Sabir, A., & Sikandar, S. (2021). *Trichoderma*: Biodiversity, abundances, and biotechnological applications. *Recent Trends in Mycological Research: Volume 1: Agricultural and Medical Perspective*, 293-315.
- Alshelmani, M. I., Abdalla, E. A., Kaka, U., & Basit, M. A. (2021). Nontraditional feedstuffs as an alternative in poultry feed. In *Advances in Poultry Nutrition Research*. IntechOpen.
- Alshelmani, M. I., Loh, T. C., Foo, H. L., Sazili, A. Q., & Lau, W. H. (2017). Effect of feeding different levels of palm kernel cake fermented by *Paenibacillus polymyxa* ATCC 842 on broiler growth performance, blood biochemistry, carcass characteristics, and meat quality. *Animal Production Science*, 57(5), 839–848. <https://doi.org/10.1071/AN15359>
- Ashayerizadeh, A., Dastar, B., Shargh, M. S., Mahoonak, A. R. S., & Zerehdaran, S. (2018). Effects of feeding fermented rapeseed meal on growth performance, gastrointestinal microflora population, blood metabolites, meat quality, and lipid metabolism in broiler chickens. *Livestock Science*, 216, 183–190. <https://doi.org/10.1016/j.livsci.2018.08.012>
- Bayitse, R., Hou, X., Laryea, G., & Bjerre, A.-B. (2015). Protein enrichment of cassava residue using *Trichoderma pseudokoningii* (ATCC 26801). *AMB Express*, 5, 80. <https://doi.org/10.1186/s13568-015-0166-8>
- Chiang, G., Lu, W. Q., Piao, X. S., Hu, J. K., Gong, L. M., & Thacker, P. A. (2009). Effects of feeding solid-state fermented rapeseed meal on performance, nutrient digestibility, intestinal ecology and intestinal morphology of broiler chickens. *Asian-Australasian Journal of Animal Sciences*, 23(2), 263-271.
- Choudhary, M., Sharma, P. C., Jat, H. S., Nehra, V., McDonald, A. J., & Garg, N. (2016). Crop residue degradation by fungi isolated from conservation agriculture fields under rice–wheat system of North-West India. *International Journal of Recycling of Organic Waste in Agriculture*, 5, 349-360.
- Chu, Y. T., Lo, C. T., Chang, S. C., & Lee, T. T. (2017). Effects of *Trichoderma* fermented wheat bran on growth performance, intestinal morphology and histological findings in broiler chickens. *Italian Journal of Animal Science*, 16(1), 82–92. <https://doi.org/10.1080/1828051X.2016.1241133>
- Chuang, W. Y., Lin, W. C., Hsieh, Y. C., Huang, C. M., Chang, S. C., & Lee, T. T. (2019). Evaluation of the combined use of *Saccharomyces cerevisiae* and *Aspergillus oryzae* with phytase fermentation products on growth, inflammatory, and intestinal morphology in broilers. *Animals*, 9(12), 1–16. <https://doi.org/10.3390/ani9121051>
- Elmasry, M., Elgreimi, S. M., Belal, E., Elmostafa, K. E., & Eid, Y. (2017). Assessment of The Performance of Chicks Fed with Wheat Bran Solid Fermented by *Trichoderma longibrachiatum* (SF1). *Journal of Sustainable Agricultural Sciences*, 43(2), 115–126. <https://doi.org/10.21608/jsas.2017.1162.1008>
- Ezekiel, O. O., & Aworh, O. C. (2013). Solid State Fermentation of Cassava Peel with Enrichment. *International Journal of Biological, Food, Veterinary and Agricultural Engineering*, 7(3), 202–209.
- Gungor, E., & Erener, G. (2020). Effect of dietary raw and fermented sour cherry kernel (*Prunus cerasus* L.) on growth performance, carcass traits, and meat quality in broiler chickens.

- Poultry Science, 99(1), 301–309. <https://doi.org/10.3382/ps/pez490>
- Hatta U, Sjoifjan O, S. I. and S. B. (2014). Effects of fermentation by *Trichoderma viride* on nutritive value of copra meal, cellulase activity and performance of broiler chickens. *Livestock Research for Rural Development*. <https://lrrd.cipav.org.co/lrrd26/4/hatt26061.htm>
- Ikpe, J. N., Oko, E. C., & Vining-Ogu, I. C. (2019). Potentials of Bio Fermented Rice Husk Meal as a Replacement to Brewer's Dried Grain in Finisher Broiler's Diet. *Journal of Agricultural Science (Toronto)*, 11(2), 533-540. <https://doi.org/10.5539/jas.v11n2p533>.
- Jangir, M., Pathak, R., & Sharma, S. (2017). *Trichoderma* and its potential applications. *Plant-Microbe Interactions in Agro-Ecological Perspectives: Volume 2: Microbial Interactions and Agro-Ecological Impacts*, 323-339. [https://doi.org/10.1007/978-981-10-6593-4\\_13](https://doi.org/10.1007/978-981-10-6593-4_13).
- Junejo, S. A., Zhang, L., Yang, L., Wang, N., Zhou, Y., Xia, Y., & Wang, H. (2019). Anti-hyperlipidemic and hepatoprotective properties of wheat bran with different particle sizes. *Journal of the Science of Food and Agriculture*, 99(4), 1990-1996. <https://doi.org/10.1002/jsfa.9397>
- Lee, S. K., Chon, J. W., Yun, Y. K., Lee, J. C., Jo, C., Song, K. Y., Kim, D. H., Bae, D., Kim, H., Moon, J. S., & Seo, K. H. (2022). Properties of broiler breast meat with pale color and a new approach for evaluating meat freshness in poultry processing plants. *Poultry Science*, 101(3), 101627. <https://doi.org/10.1016/j.psj.2021.101627>
- Lin, W. C., Lee, M. T., Lo, C. T., Chang, S. C., & Lee, T. T. (2018). Effects of dietary supplementation of *Trichoderma pseudokoningii* fermented enzyme powder on growth performance, intestinal morphology, microflora and serum antioxidative status in broiler chickens. *Italian Journal of Animal Science*, 17(1), 153–164. <https://doi.org/10.1080/1828051X.2017.1355273>
- Marcinčák, S., Klempová, T., Bartkovský, M., Marcinčáková, D., Zdolec, N., Popelka, P., Mačanga, J., & Čertík, M. (2018). Effect of fungal solid-state fermented product in broiler chicken nutrition on quality and safety of produced breast meat. *BioMed Research International*, 2018. <https://doi.org/10.1155/2018/2609548>
- Mondal, S. (2021). Tests for specific carbohydrates : Anthrone test ; Mucic acid test ; Osazone test ; Test for non-reducing sugars ; Bial ' s test . Anthrone Test Principle Uses of Anthrone Test Limitations of Anthrone Test Mucic acid test Principle Mucic acid Test Procedu. November.
- Oguri M, Okano K, Ieki H, Kitagawa M, Tadokoro O, Sano Y, Oishi K, Hirooka H, Kumagai H. 2013. Feed intake, digestibility, nitrogen utilization, ruminal condition and blood metabolites in wethers fed ground bamboo pellets cultured with white-rot fungus (*Ceriporiopsis subvermispora*) and mixed with soybean curd residue and soy sauce cake. *Anim Sci J*. 84:650–655.
- Omwango, E. O., Njagi, E. N. M., Orinda, G. O., & Wanjau, R. N. (2013). Nutrient Enrichment of Pineapple Waste using *Aspergillus niger* and *Trichoderma viride* by Solid State Fermentation. *African Journal of Biotechnology*, 12(43), 6193–6196. <https://doi.org/10.5897/ajb2013.12992>
- Predescu, N. C., Stefan, G., Rosu, M. P., & Papuc, C. (2024). Fermented Feed in Broiler Diets Reduces the Antinutritional Factors, Improves Productive Performances and Modulates Gut Microbiome—A Review. *Agriculture*, 14(10), 1752.
- Saini, P., Islam, M., Das, R., Shekhar, S., Sinha, A. S. K., & Prasad, K. (2023). Wheat bran as potential source of dietary fiber: Prospects and challenges. *Journal of Food Composition and Analysis*, 116, 105030.
- Satpathy, L., Dash, D., Sahoo, P., Anwar, T. N., & Parida, S. P. (2020). Quantitation of Total Protein Content in Some Common Edible Food Sources by Lowry Protein Assay. *Letters in Applied NanoBioScience*, 9(3), 1275–1283. <https://doi.org/10.33263/lianbs93.12751283>

- Sharma, R. K., & Arora, D. S. (2011). Solid state degradation of paddy straw by *Phlebia floridensis* in the presence of different supplements for improving its nutritive status. *International Biodeterioration & Biodegradation*, 65(7), 990–996. <https://doi.org/10.1016/J.IBIDOD.2011.07.007>
- Srivastava, N., Srivastava, M., Ramteke, P. W., & Mishra, P. K. (2019). Solid-state fermentation strategy for microbial metabolites production: An overview. In *New and Future Developments in Microbial Biotechnology and Bioengineering: Microbial Secondary Metabolites Biochemistry and Applications*. Elsevier B.V. <https://doi.org/10.1016/B978-0-444-63504-4.00023-2>
- Sugiharto, S., & Ranjitkar, S. (2019). Recent advances in fermented feeds towards improved broiler chicken performance , gastrointestinal tract microecology and immune responses: A review. *Animal Nutrition*, 5(1), 1–10. <https://doi.org/10.1016/j.aninu.2018.11.001>
- Sugiharto, S., Yudiarti, T., Isroli, I., Widiastuti, E., & Putra, F. D. (2017). Effects of feeding cassava pulp fermented with *Acremonium charticola* on growth performance, nutrient digestibility and meat quality of broiler chicks. *South African Journal of Animal Sciences*, 47(2), 130–139. <https://doi.org/10.4314/SAJAS.V47I2.4>
- Teng, P. Y., Chang, C. L., Huang, C. M., Chang, S. C., & Lee, T. T. (2017). Effects of solid-state fermented wheat bran by *Bacillus amyloliquefaciens* and *Saccharomyces cerevisiae* on growth performance and intestinal microbiota in broiler chickens. *Italian Journal of Animal Science*, 16(4), 552-562.

**Table 1. Protein content of fermented and unfermented substrates**

Substrate samples	Protein concentration (mg/g)	P-value
Unfermented wheat bran	19.06±0.13 mg	0.000
<i>Trichoderma asperellum</i> fermented wheat bran	54.51±0.09 mg	
Unfermented rice husk	10.56±0.06 mg	0.049
<i>Trichoderma asperellum</i> fermented rice husk	10.80±0.06 mg	

Values symbolize the average (mean) of 3 replicates of each treatment group along with standard error.

**Table 2. Carbohydrate content of fermented and unfermented substrates**

Substrate samples	Carbohydrate concentration (mg/g)	P-value
Unfermented wheat bran	16.45±0.07 mg	0.000
<i>Trichoderma asperellum</i> fermented wheat bran	22.58±0.14 mg	
Unfermented rice husk	6.45±0.06 mg	0.718
<i>Trichoderma asperellum</i> fermented rice husk	6.50±0.12 mg	

Values symbolize the average (mean) of 3 replicates of each treatment group along with standard error.

**Table 3. Effect of designed feed mixtures on average weekly live body weight of broiler chickens**

	DESIGNED FEED MIXTURES					P-value
	T1	T2	T3	T4	T5	
Weeks	Control	Diet with 10% WB	Diet with 10% FWB	Diet with 10% RH	Diet with 10% FRH	
week 0	43.14±0.51	43.60±0.42	43.54±0.48	43.34±0.54	43.56±0.59	0.965
week 1	138.8±3.60	134±1.70	135.6±1.50	131.6±2.48	130.6±3.76	0.317
week 2	337.4 <sup>ab</sup> ±5.89	323.2 <sup>b</sup> ±14.0	370.8 <sup>a</sup> ±14.7	306.2 <sup>b</sup> ±16.7	310.6 <sup>b</sup> ±7.15	0.011
week 3	618 <sup>b</sup> ±8.60	616.4 <sup>b</sup> ±11.7	672 <sup>a</sup> ±8.60	606.2 <sup>b</sup> ±19.0	614.2 <sup>b</sup> ±13.3	0.012
week 4	900.4 <sup>b</sup> ±21.7	943.4 <sup>ab</sup> ±22.4	989.2 <sup>a</sup> ±11.7	931.4 <sup>b</sup> ±11.8	928.4 <sup>b</sup> ±11.4	0.019
week 5	1304 <sup>b</sup> ±23.3	1296.2 <sup>b</sup> ±17.3	1380 <sup>a</sup> ±11.4	1293.4 <sup>b</sup> ±16.5	1311.8 <sup>b</sup> ±15.1	0.011

Values symbolize the average (mean) of 5 models of each treatment group along with standard error. Means within same rows with different superscript letters (ab) are significantly different ( $p < 0.05$ ). WB: wheat bran, FWB: fermented wheat bran, RH: rice husk, FRH: fermented rice husk.

### Body weight week 1 to 5

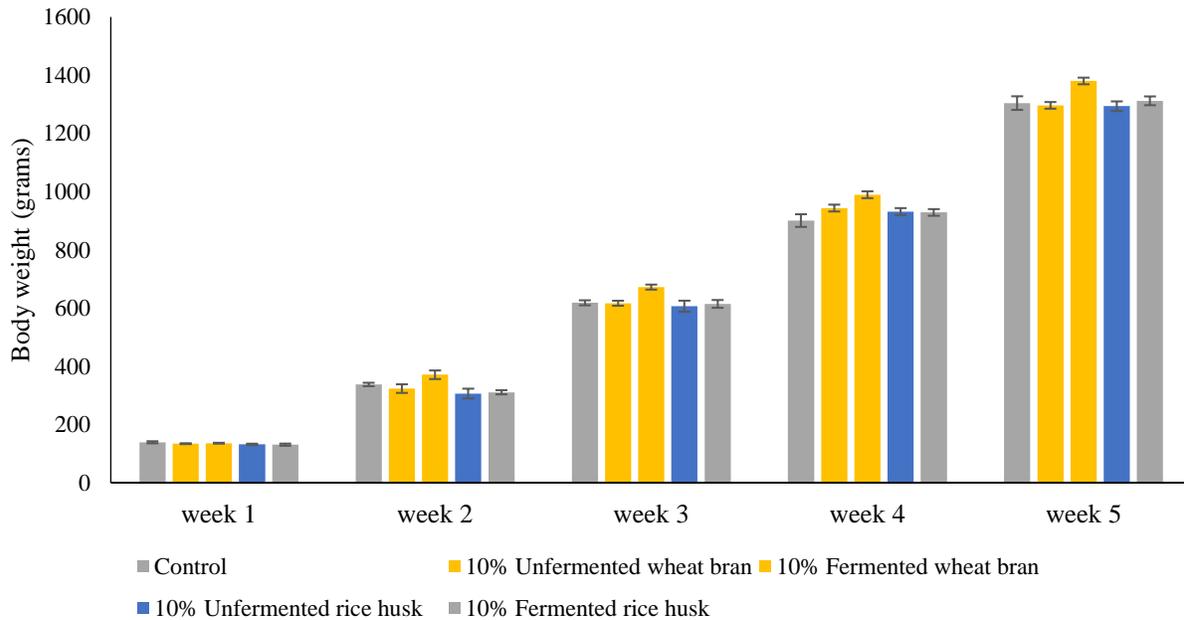


Figure 1. Effect of designed feed mixtures on the body weight of broiler chicks during the experiment study Each bar represents average body weight along with standard error (n=5). Fermented wheat bran-based feed improved the body weight in each successive week shown with yellow bar.

**Table 4: Effect of designed feed mixtures on breast meat colour and pH of broiler chicken**

Parameters	DESIGNED FEED MIXTURES						P-value
	Control	WB	FWB	RH	FRH	SEM	
Breast pH	5.82	5.71	5.77	5.58	5.71	0.021	0.261
L* (Lightness)	56.97	57.22	57.36	56.29	58.39	0.239	0.083
a* (redness)	3.74	3.78	4.09	3.94	3.76	0.120	0.890
b* (yellowness)	6.49	6.42	6.84	7.26	6.54	0.112	0.092

Values symbolize the average (mean) of 5 models of each treatment group. WB: wheat bran, FWB: fermented wheat bran, RH: rice husk, FRH: fermented rice husk. SEM: standard error of mean.

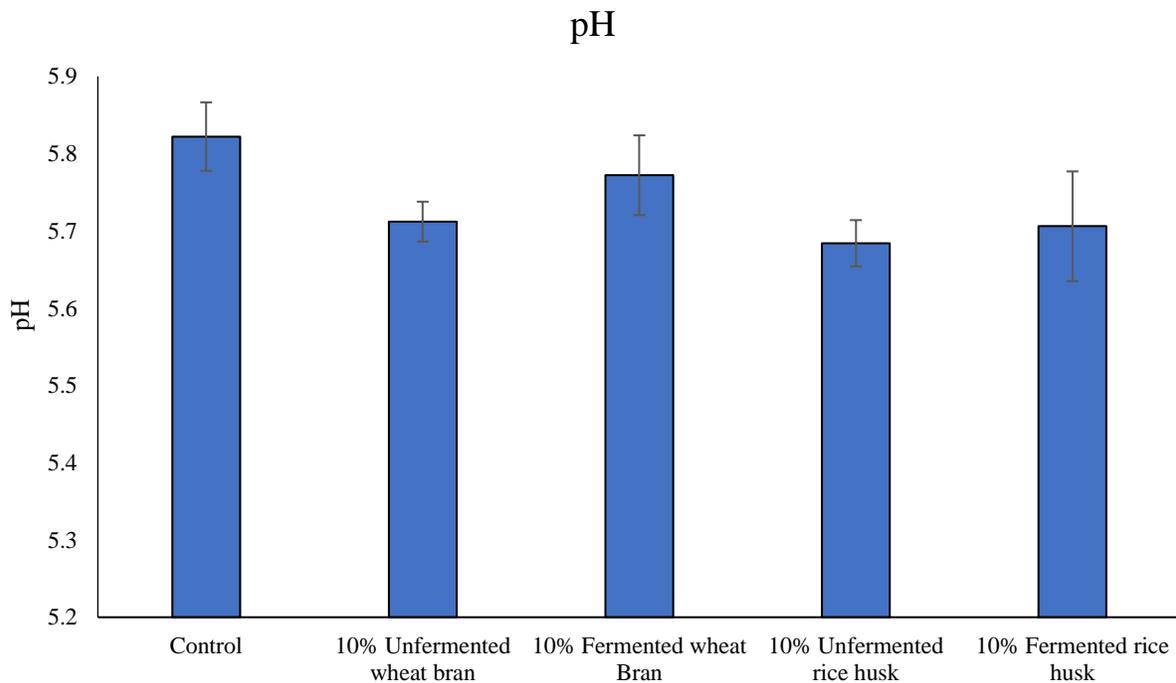


Figure 2. Effect of experimental diets on the pH of broiler chicken. Each bar represents average pH along with standard error (n=5). No significant difference was detected between treatment groups (Least significant difference,  $P < 0.05$ ).

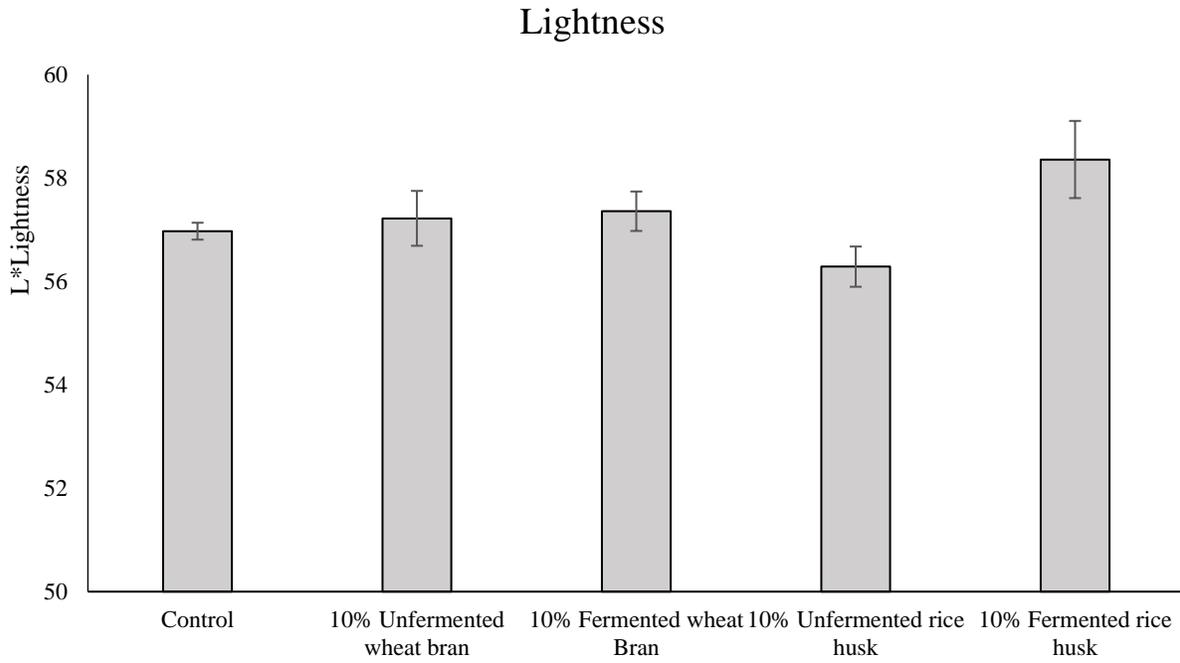


Figure 3. Effect of experimental diets on the breast meat colour (L\*) of broiler chicken. Each bar represents average values of lightness along with standard error (n=5). No significant difference was detected between treatment groups (Least significant difference, P<0.05).